

trans-2,6-, 3,6- AND 4,6-DIAZA-5,6,6a,7,8,12b-HEXAHYDRO-BENZO[C|PHENANTHRENE-10,11-DIOLS AS DOPAMINE AGONISTS

Yu Gui Gu,* Erol K. Bayburt, Michael R. Michaelides, Chun Wel Lin, and Kazumi Shiosaki

Neuroscience Research, Pharmaceutical Discovery, Abbott Laboratories, 100 Abbott Park Road, Abbott Park, IL 60064-3500, U.S.A.

Received 17 September 1998; accepted 9 November 1998

Abstract: The title compounds were synthesized by replacing the thiophene moiety of A-86929(2a) with variously substituted pyridines. Dopamine D-1 and D-2 binding and adenylate cyclase assays indicate that 4,6-diaza compounds 15 are potent and selective full D1 agonists when R^1 is H or a small substituent and $R^2 = H$, with D1 binding affinity and adenylate cyclase functional potency equivalent to that of A-86929(2a). © 1999 Elsevier Science Ltd. All rights reserved.

Dopamine is an important neurotransmitter in the central nervous system (CNS). Dopamine imbalance is believed to play a key role in a number of CNS-related disorders such as schizophrenia, Parkinson's disease (PD), drug abuse, eating disorders, and depression. Dopamine also has several important roles in the peripheral nervous system, such as in the control of blood to the kidneys and in autonomic ganglion transmission. Dopamine receptors have traditionally been divided into two general receptor classes, designated D-1-like and D-2-like, based on biochemical and pharmacological differences between the two receptor types. Recent molecular biology studies have indicated an even greater heterogeneity of dopamine receptors: the D-2, D-3, and D-4 constitute the D-2-like class, while the D-1 and D-5 receptors comprise the D-1-like class.²

PD is a progressive disease that arises as a result of the degeneration of dopaminergic neurons in the zona compacta of the substantia nigra.³ Currently, levodopa remains the most effective treatment for PD.⁴ However, significant loss of efficacy and severe side effects⁵ associated with the chronic treatment of levodopa warrant the development of alternative therapies. Dopamine D2 agonists have been used for the treatment of PD, often in combination with levodopa.⁶

Recently, the development of dopamine D1 agonists as an alternative for the treatment of PD emerged with the hope of improving the side effect profile. Dihydrexidine(1),⁷ a selective D1 full agonist, showed a promising preclinical pharmacological profile. A-86929(2a) is also a selective full D1 agonist that is effective in rodent and primate models of PD after both acute and chronic administration.⁸ ABT-431 (2b)⁸, the diacetyl prodrug of A-86929(2a), has shown efficacy in human clinical trials.⁹ We have extended our SAR studies by replacing the thiophene moiety of A-86929(2a) with variously substituted pyridines and would like to report our latest results.

Synthesis

Treatment of t-butyl 2-(or 4-)pyridinecarboxamide derivatives 4^{10} (or 5^{11}) with 2 equiv of n-BuLi at -78 °C generated the 3-lithiopyridine dianions which were condensed with nitroolefin 6^8 followed by the

triethylamine-mediated equilibration of the resulting *cis/trans* adduct mixture to afford compounds 7 and 8 in low yield. Reduction of the nitro group of compounds 7 and 8 with Zn/HCl provided the corresponding primary amines 9 and 10. When compounds 7 and 8 were reacted with large excess of zinc dust and 6 N HCl, in addition to the formation of the primary amines 9 and 10, the tetracyclic compounds 13 and 14 were obtained directly in poor to moderate yield, accompanied by the formation of the diamines with both nitro and amide groups being reduced. Further treatment of 9 and 10 with excess of Zn/6 N HCl did not provide any cyclized compounds 13 and 14, indicating that 9 and 10 are not intermediates involved in the formation of compounds 13 and 14. When 7 ($R^1 = F$, $R^2 = H$) was reacted with a large excess of zinc dust and 6 N HCl, however, the tetracyclic compound 13 ($R^1 = F$, $R^2 = H$) was not observed. Compound 13 ($R^1 = F$, $R^2 = H$) was synthesized in an alternative three-step sequence. Reduction of nitro group to an amine 9 ($R^1 = F$, $R^2 = H$) followed by treatment with acid in refluxing toluene provided the lactam 11 ($R^1 = F$, $R^2 = H$) in 60% yield. Reduction of the lactam 11 ($R^1 = F$, $R^2 = H$) with borane afforded 13 ($R^1 = F$, $R^2 = H$) and the desfluoro compound (13, $R^1 = R^2 = H$) in 23% and 18% yield, respectively (Scheme 1). The cleavage of the methyl ethers of 13 and 14 with boron tribromide finally provided catecholamines 15 and 16.

In contrast, when $5 (R^1 = H, R^2 = Me)$ was treated with 2 equiv of *n*-BuLi and then reacted with the nitroolefin 6, the desired Michael adduct $8 (R^1 = H, R^2 = Me)$ was not observed. Instead, compound 17 was isolated as the major product via lithiation of the methyl hydrogen¹² of $5 (R^1 = H, R^2 = Me)$ and subsequent addition of the resulting anion to nitroolefin 6. An alternative synthetic route was developed to synthesize the target compound 16c (Scheme 2). Halogen-metal exchange of 3-bromo-5-methyl-4-pyridinemethanol MOM ether 18^{13} with *n*-BuLi followed by addition of the 3-lithiopyridine intermediate to nitroolefin 6^8 and subsequent base-promoted equilibration gave rise to compound 20 in good yield. The reduction of the nitro group with 2n/6 N HCl, cleavage of MOM ether, and protection of the primary amine afforded 22 in good yield. The construction of the piperidine ring was accomplished via a three-step sequence: Conversion of the primary alcohol 22 to the corresponding chloride, deprotection of the amine, and ring closure provided compound 23. Finally, cleavage of methyl ether with boron tribromide afforded target molecule 16c.

Compound 26, a regioisomer of compound 16, was synthesized from 4-bromo-3-pyridinemethanol benzyl ether 19¹⁴ by employing a similar synthetic route (Scheme 2). The overall yield of compound 26 is 14% from starting material 19.

Scheme 2

Results and Discussion

The binding affinities to both human D1 and human D2 receptors were measured, and the results for the 4,6-diaza derivatives are shown in Table 1. Dihydrexidine (1) and A-86929(2a) are included as reference compounds. In the 4,6-diaza series, the unsubstituted pyridine compound 15a ($R^1 = R^2 = H$) exhibited high affinity for the D1 receptor ($K_i = 65$ nM). 15a has high potency in adenylate cyclase functional assay ($EC_{50} = 6.7$ nM) and greater intrinsic activity relative to dopamine (IA = 125%). These results indicate that 15a is a potent, full agonist at the D1 receptor, with an in vitro profile comparable to that produced by A-86929(2a). Compound 15a has significantly weaker affinity for the D2 receptor ($K_i = 2,500$ nM), thus a more selective D1 agonist (D2/D1 = 38) than A-86929 (2a) (D2/D1 = 14).

Previous structure–activity relationship (SAR) studies on D1 ligands suggested that the D1 but not the D2 receptor possesses a lipophilic binding pocket that can bind a bulky substituent such as adamantyl group. ¹⁵ SAR studies on A-86929 analogs (27) and its regioisomers(28) revealed that introduction of a small R group (methyl, ethyl or n-propyl) increases affinity for D1-like receptor relative to the parent compounds (27 and 28, R = H). Increase in D2 receptor affinity by 27 and 28 relative to the parent was not as significant. ¹⁶ Nichols and coworkers

reported that dihydrexidine analog 29 (R = Me) did not produce such a boost in affinity relative to the parent.¹⁷ In contrast, introduction of a methyl group at C3 position of the 4,6-diaza series (15b) resulted in a 2-fold decrease in D1 affinity and 2-fold increase in D2 affinity, and therefore a 4-fold drop of D1 selectivity relative to the parent compound 15a. Interestingly, C3 fluorine substituted compound 15c showed similar D1 binding affinity and selectivity to 15b, indicating that the contribution of electronic effect to dopamine affinity is relatively insignificant in the 4,6-diaza series. When a larger substituent is introduced at the C3 position (15d,e), D1 affinity remains unaffected, whereas D2 affinity increases by about 4-fold, leading to a lower D2/D1 ratio compared to the parent compound 15a. It is interesting to note that when the size of the substituent R¹ increases, D2 affinity increases accordingly (n-Pr > OMe > Me > F > H), whereas the effect on D1 affinity varies. In an adenylate cyclase based functional assay, compounds 15d,e exhibited high potency as the parent compound 15a, while 15b,c were less potent (2- to 3-fold) than 15a, in parallel with their decreased D1 binding affinity.

Table 1. Competitive binding and D1 functional activities of 4,6-diaza-5,6,6a,7,8,12b-hexahydrobenzo[c]phenanthrene-10.11-diols*

compd	R¹	R²	D1			D2	***
			K _i (nM)	EC ₅₀ (nM)	IA (%) ^b	K _i (nM)	D2/D1
1			$33 \pm 2(6)$	14 ± 5(6)	121 ± 7(6)	$1520 \pm 527(5)$	45
2a			$49 \pm 5(32)$	$9 \pm 2(29)$	$125 \pm 3(29)$	$710 \pm 80(52)$	14
15a	Н	Н	$65 \pm 7(6)$	$6.7 \pm 2(3)$	$125 \pm 3(3)$	$2500 \pm 620(5)$	38
15b	Me	H	$140 \pm 14(10)$	$23 \pm 4(3)$	$121 \pm 6(3)$	$1400 \pm 390(9)$	10
15c	F	H	$180 \pm 16(5)$	$18 \pm 3(3)$	$141 \pm 13(3)$	$2200 \pm 500(3)$	12
15d	OMe	Н	$49 \pm 11(6)$	$8.8 \pm 2(3)$	$114 \pm 13(3)$	$520 \pm 72(5)$	11
15e	n-Pr	Н	$45 \pm 5(5)$	$8.6 \pm 2(3)$	$116 \pm 12(3)$	$450 \pm 84(6)$	10
15f	Н	Me	$240 \pm 43(6)$	$110 \pm 41(4)$	$156 \pm 4(4)$	12000(1)	50
15g	H	n-Bu	$890 \pm 230(7)$	$180 \pm 58(3)$	$118 \pm 7(3)$	$1900 \pm 220(6)$	2
15h	Me	Me	$280 \pm 37(6)$	$120 \pm 44(3)$	$134 \pm 11(3)$	9100 ± 1500(3)	33
15i	-(CH ₂) ₄ -		$560 \pm 68(6)$	$63 \pm 5(3)$	$136 \pm 9(3)$	3600(1)	6

^aValues represent the mean ±SEM, with the number of experiments in parentheses. Binding and adenylate assays were carried out in HEK and LTK cells transfected with the human D1¹⁸ and D2¹⁹ receptors, respectively. Binding ligands were as follows: D1, [¹²⁵I]SCH23982; D2, [³H]-spiperone. Binding K_i and cyclase EC₅₀ values were determined as previously described. ²⁰ bIA = Intrinsic activity relative to dopamine.

A significant loss (4-fold) of D1 affinity was observed when the parent compound 15a was substituted with a methyl group at the C2 position (15f). In contrast to compound 15b, 15f also showed a 5-fold decrease of D2 binding affinity, thus a higher D1 selectivity (D2/D1 = 50) than 15a. Previous studies on dihydrexidine analogs 30 also indicated that the optimal D2/D1 selectivity was achieved when $R = Me.^{17}$ Increasing the size of the substituent at C2 position (e.g., $R^2 = n$ -Bu) resulted in a further decrease in D1 binding affinity and increase in D2 affinity (15g vs 15f). C2 and C3 disubstituted compounds also showed a decreased D1 binding affinity relative to the parent compound 15a. Dimethyl substituted compound 15h is four times less potent than 15a in both D1 and D2 binding. Further increasing the size of the substituents at C2 and C3 positions resulted in a decrease in D1 binding affinity but an increase in D2 affinity (15h vs 15i). In adenylate cyclase based functional assay, C2 monosubstituted (15f,g) and C2,C3 disubstituted (15h,i) 4,6-diaza derivatives showed lower potency relative to the parent compound 15a and their C3 monosubstituted analogs (15b-e). Although the same trend is

also observed in D1 binding assay as described above, the decrease of functional activity (10- to 27-fold) is more dramatic than that of D1 binding affinity (4- to 14-fold). 4,6-Diaza derivatives have greater intrinsic activity relative to dopamine.

Previous studies revealed that the different regioisomers of thiophene derivatives 27, 28, and 31 exhibited comparable affinity for the D1 receptor, although 28 showed higher (ca. 3-fold) affinity on D2 binding than the other two regioisomers. In the current study, D1 affinity decreases dramatically (17-fold) when the pyridine nitrogen is moved from C4 to C2 position (15a vs 16a), whereas the effect on D2 binding is less pronounced, with a 2-fold increase in D2 binding affinity (Table 2). An even more dramatic decrease in functional activity (48-fold) was observed. This result may provide insight for the design of selective D1 ligands. The reasons for the significant decrease of D1 affinity of 16a remain unclear, although may be due to the unfavorable electrostatic interaction of the lone pair electrons of the pyridine nitrogen with the D1 receptor. When a strong electron-withdrawing substituent was introduced to reduce the availability of the lone pair electrons, an increase (3-fold) in D1 binding affinity was observed (16b). The methyl substituted analog 16c showed the same D1 binding affinity and a 3-fold increase in D2 binding affinity relative to its parent compound 16a. In fact, compound 16c is the only compound in this study which binds more potently to the D2 receptor than the D1 receptor in dopamine binding assay. The 2,6-Diaza derivatives 16a-c also exhibited weak potency in stimulating adenylate cyclase (EC₅₀ = 0.23-0.80 μM). It is noteworthy that compound 16b is 2.5- to 3.5-fold less potent in D1 functional assay than 16a and 16c, although 16b showed higher (3-fold) D1 binding affinity.

Compound 26, the only compound in the 3,6-diaza series tested in this study, exhibited weaker dopamine binding affinity (2-fold) and functional activity (6-fold) relative to its regioisomer 15a.

Table 2. Competitive binding and D1 functional activities of 2,6- and 3,6-diaza-5,6,6a,7,8,12b-hexahydrobenzo-[c]phenanthrene-10,11-diols*

			D2			
compd	\mathbb{R}^2	K _i (nM)	EC ₅₀ (nM)	IA (%) ^b	K, (nM)	D2/D1
2a	-	$49 \pm 5(32)$	9 ± 2(29)	125 ± 3(29)	$710 \pm 80(52)$	14
16a	H	$1100 \pm 310(8)$	$320 \pm 150(3)$	$81 \pm 8(3)$	$1300 \pm 480(8)$	1
16b	F	$390 \pm 110(3)$	$800 \pm 230(3)$	$126 \pm 11(3)$	890 ± 170(7)	2
16c	Me	$1100 \pm 160(3)$	$230 \pm 59(4)$	$75 \pm 12(4)$	$400 \pm 100(3)$	0.36
26		$140 \pm 20(6)$	$38 \pm 17(4)$	$131 \pm 7(4)$	$4400 \pm 1400(6)$	31

^aValues represent the mean ±SEM, with the number of experiments in parentheses. Binding and adenylate assays were carried out in HEK and LTK cells transfected with the human D1¹⁸ and D2¹⁹ receptors, respectively. Binding ligands were as follows: D1, [¹²³I]SCH23982; D2, [³H]-spiperone. Binding K, and cyclase EC₅₀ values were determined as previously described. ²⁰ bIA = Intrinsic activity relative to dopamine.

In conclusion, different pyridine regioisomers of A-86929(2a) have been synthesized, and their binding affinity to D1 and D2 receptors and functional activity in adenylate cyclase assay have been determined. In the 2,6-diaza series, when R^2 is H and R^1 is H or a small substituent, compounds 15a—e are full D1 agonists (>100% intrinsic activity relative to dopamine), exhibiting high D1 binding affinity and selectivity (D2/D1 \geq 10). Substitution at the C2 position (15, $R \neq H$) resulted in a decreased D1 binding affinity and functional activity, whereas effect on D2 binding varies (15f–i). 4,6-Diaza derivatives 16 are poor dopamine ligands, showing low affinity and poor selectivity for dopamine receptors.

Acknowledgment: We would like to thank Bruce Bianchi, Tom Miller, Ellen Roberts and Michael Stashko for their technical assistance.

References and Notes

- (a) Kebabian, J. W.; Calne, D. B. Nature (London) 1979, 277, 93. (b) For a review, see Kaiser, C.; Jain, T. Med. Res. Rev. 1985, 5, 145.
- 2. Sibley, D. R.; Monsma, F. J., Jr. Trends Pharmacol. Sci. 1992, 13, 61.
- 3. Agid, Y. Lancet 1991, 337, 1321.
- 4. Cotzias, G. C.; Van-Woert, M. H.; Schiffer, L. M. N. Engl. J. Med. 1967, 276, 374.
- 5. Sage, J. I.; Mark, M. H. Neurology 1994, 44(suppl 6), S10-S14.
- 6. Standaert, D. G.; Stern, M. B. Medical Clinics of North America 1993, 77(1), 169.
- (a) Brewster, W. K.; Nichols, D. E.; Riggs, R. M.; Mottola, D. M.; Lovenberg, T. W.; Lewis, M. H.; Mailman, R. B. J. Med. Chem. 1990, 33, 1756.
 (b) Knoerzer, T. A.; Nichols, D. E.; Brewster, W. K.; Watts, V. J.; Mottola, D.; Mailman, R.B. J. Med. Chem. 1994, 37, 2453.
- 8. Michaelides, M. R.; Hong, Y.; DiDomenico, S., Jr.; Asin, K. E.; Britton, D. R.; Lin, C. W.; Williams, M.; Shiosaki, K; J. Med. Chem. 1995, 38, 3445.
- Brefel, C.; Blin, O.; Descombes, S.; Soubrouillard, C.; Fabre, N.; Viallet, F.; Thalamas, C.; Azulay, J. P.; Senard, J. M.; Montastruc, J. L.; Lafnitzegger, K.; Frederick, E.; Wright, S.; Nutt, J. G.; Rascol, O. 49th Annual Meeting, American Academy of Neurology, Boston, April 1997.
- 10. Prepared from the corresponding pyridinecarboxylic acids unless otherwise indicated. The pyridinecarboxylic acids are either commercially available (R¹ = R² = H; R¹ = Me, R² = H; R¹ = H, R² = n-Bu) or synthesized according to Shuman et al., J. Org. Chem. 1990, 55, 738 (R¹ = n-Pr, R² = H; R¹ = R² = Me; R¹ = R² = -(CH₂)₄-) and Cooper et al., Synthesis 1971, 31 (R¹ = F, R² = H). 4 (R¹ = OMe, R² = H) was prepared from 6-bromo-2-methoxypyridine (J. Org. Chem. 1990, 55, 69.) via halogen-metal exchange and subsequent reaction with t-butyl isocyanate. 4 (R¹ = H, R² = Me) was prepared from the corresponding ethyl ester (Inorg. Syn. 1994, 4112) by employing the procedure of Weinreb et al., Tetrahedron Lett. 1977, 18, 4171.
- 11. Compound 5 (R¹ = R² = H) was prepared from isonicotinic acid. Compound 5 (R¹ = H, R² = F) was synthesized from 3-fluoropyridine via lithiation with LDA (*Tetrahedron Lett.* 1980, 21, 4137) and subsequent reaction with *tert*-butyl isocyanate.
- 12. For examples, see Marsais, F.; Breant, P.; Ginguene, A.; Queguiner, G. J. Organomet. Chem. 1981, 216, 139 and ref 13.
- 13. Gu, Y. G.; Bayburt, E. Tetrahedron Lett. 1996, 37, 2565.
- 14. Prepared from 4-bromopyridine hydrochloride via the following steps: (a) 2 equiv LDA/-78 °C then DMF/-78 °C to rt. (b) DIBAL/-78 °C. (c) NaH/BnBr/DMF/THF.
- DeNinno, M. P.; Schoenleber, R.; Perner, R. J.; Lijewski, L.; Asin, K. E.; Britton, D. R.; MacKenzie, R.; Kebabian, J. J. Med. Chem. 1991, 34, 2561.
- Michaelides, M. R.; Hong, Y.; DiDomenico, S. Jr.; Bayburt, E. K.; Asin, K. E.; Britton, D. R.; Lin, C. W.; Shiosaki, K. J. Med. Chem. 1997, 40, 1585.
- Knoerzer, T. A.; Watts, V. J.; Nichols, D. E.; Mailman, R. B. J. Med. Chem. 1995, 38, 3062.
- Lin, C. W.; Miller, T. R.; Witte, D. G.; Bianchi, B. B.; Stashko, M.; Manelli, A. M.; Frail, D. E. Mol. Pharmacol. 1995, 47, 131.
- Mouse LTH cells transfected with the human D2 receptor were provided by M. Caron at Duke University, NC.
- Kebabian, J. W.; Britton, D. R.; DeNino, M. P.; Perner, R.; Smith, L.; Jenner, P.; Schoenleber, R.; Williams, M. Eur. J. Pharmacol. 1992, 229, 203.